

# From Field Potential Waves to Ionic Current Knowledge by Leveraging Mathematical Modeling

F. Raphael<sup>2</sup>, M. Boulakia<sup>3</sup>, N. Zemzemi<sup>2</sup>, P. Zitoun<sup>1</sup>, J-F. Gerbeau<sup>2</sup>  
<sup>1</sup>Notocord, <sup>2</sup>Inria, <sup>3</sup>Sorbonne Universités UPMC

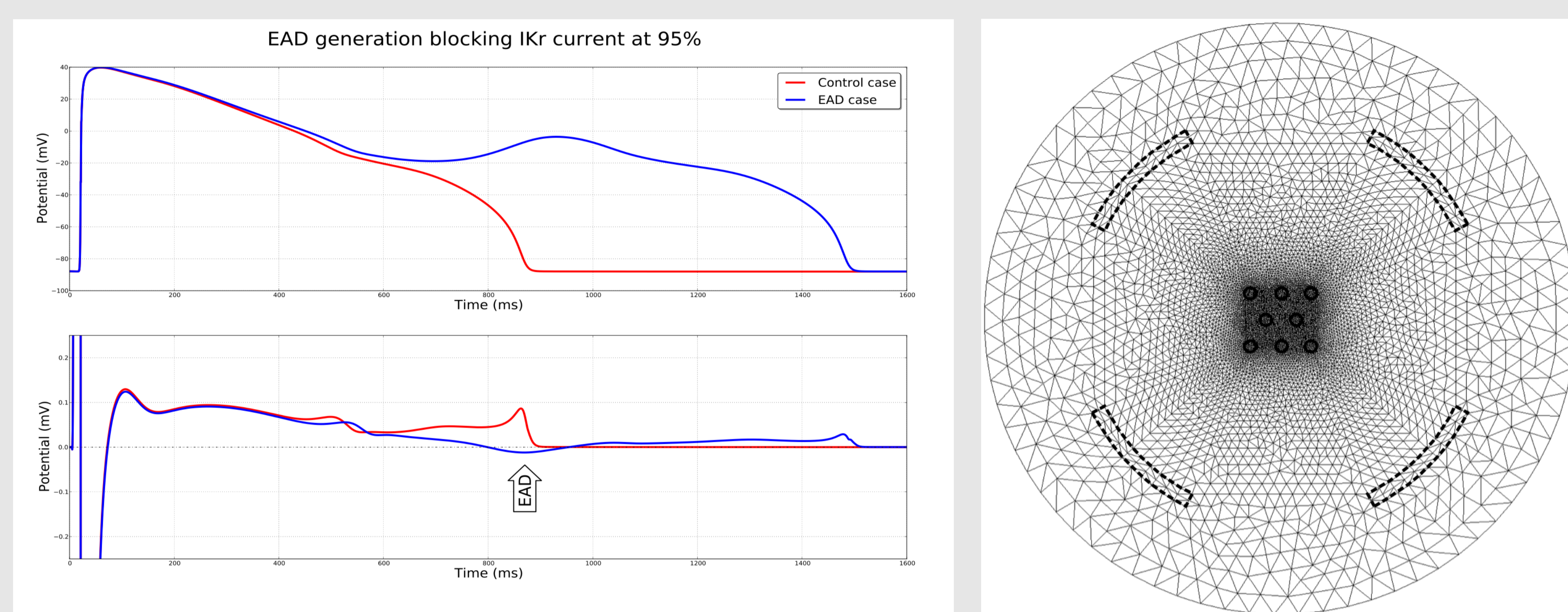
## Objectives

- The combined use of :
  - microelectrode arrays (MEAs).
  - induced pluripotent stem cells cardiomyocytes (hiPSC-CMs) allows high-throughput screening on human-derived cells.
- But the field potential (FP) signals acquired by MEAs are difficult to analyze.
- Objectives:
  - Direct problem:** propose a mathematical model of MEA signals.
  - Inverse problem:** use this model to identify channel activities.
  - Classification:** use a hybrid modeling/machine learning approach to classify compounds.

## Direct problem: concept

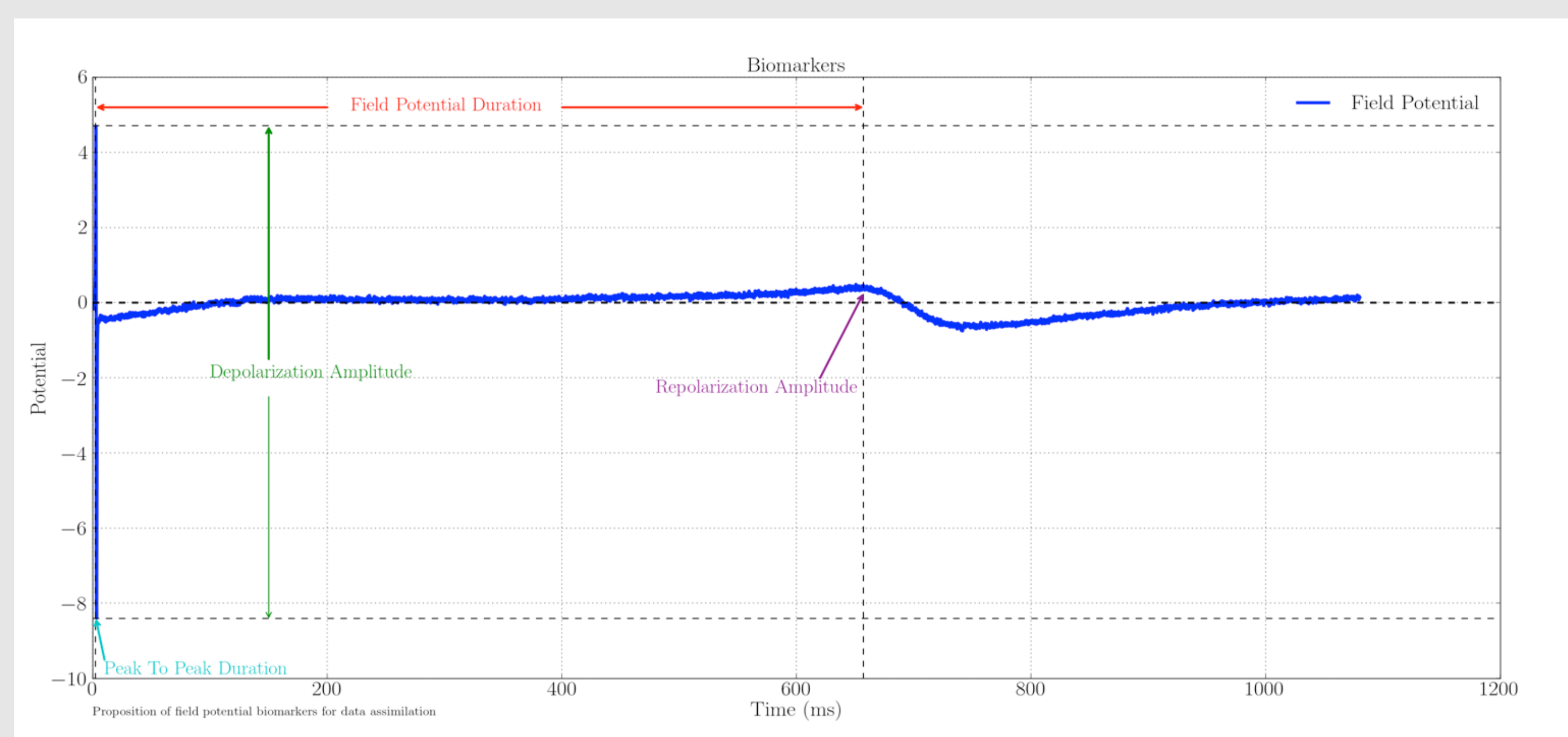
- Bidomain equations: electrophysiological model to get both transmembrane potential and extracellular potential.
- Ionic models: phenomenological (Minimal Ventricular,...) or physiological (Paci, O'Hara-Rudy,...).
- Electrode model: for the culture-electrode contact.
- Devices: well modeling.

**Example : an Early After Depolarization simulation, O'Hara-Rudy model (O'Hara et al., PLOS Comp Biol 2011), 96-well of 8 electrodes device (Axion)**



## Inverse problem: method

- Extraction of 3 biomarkers from **real signals** with NOTOCORD-fps: depolarization amplitude (DA), repolarization amplitude (RA) and field potential duration (FPD).



- Identification of channel conductance by minimizing the cost function:

$$J(\theta) = \sum_{k=1}^{N_{elec}} \left( \frac{DA_{meas}^k}{DA_{c,meas}^k} - \frac{DA^k(\theta)}{DA_c^k} \right)^2 + \left( \frac{RA_{meas}^k}{RA_{c,meas}^k} - \frac{RA^k(\theta)}{RA_c^k} \right)^2 + \left( \frac{FPD_{meas}^k}{FPD_{c,meas}^k} - \frac{FPD^k(\theta)}{FPD_c^k} \right)^2$$

$X_{meas}^k$ : experimental biomarker at electrode  $k$  (with compound)

$X^k(\theta)$ : *in silico* biomarker at electrode  $k$  with channel conductances  $\theta$

$X_{c,meas}^k$ : control experimental biomarker at electrode  $k$  (without compound)

$X_c^k$ : control *in silico* biomarker at electrode  $k$

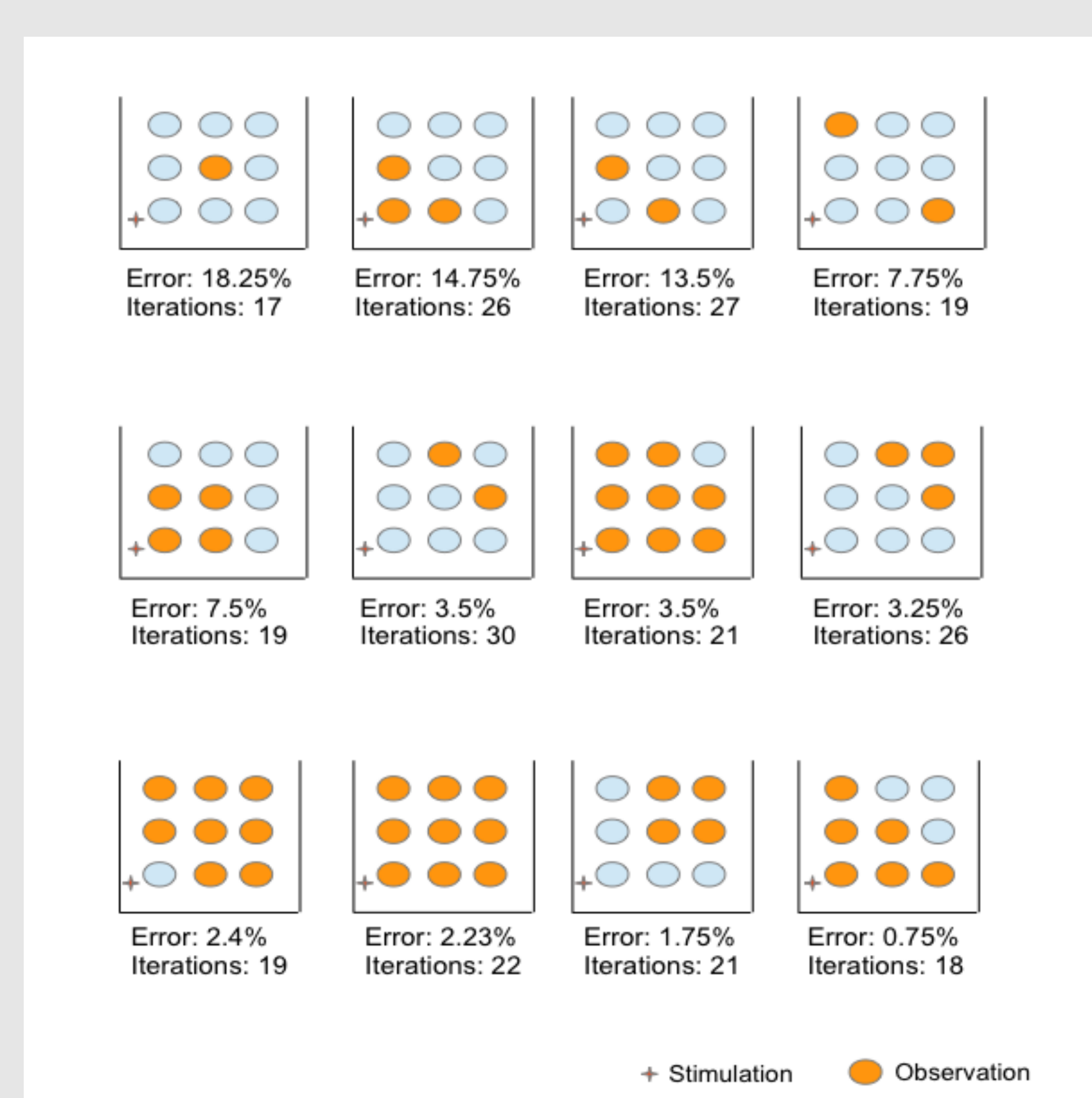
with  $X = DA, RA$  or  $FPD$

## Inverse problem: number of electrodes for the observation

- Model: MV.
- Device: 6-well of 9 electrodes (MCS).
- Data: synthetic.
- Identification of the conductance of the fast inward currents.
- By observing 1, 2, 3, 4, 6, 8 or 9 electrodes.

→ the error on the identification of the conductance depends on the relative positions of the electrodes and of the initial activation.

→ it is worth keeping all the electrodes.

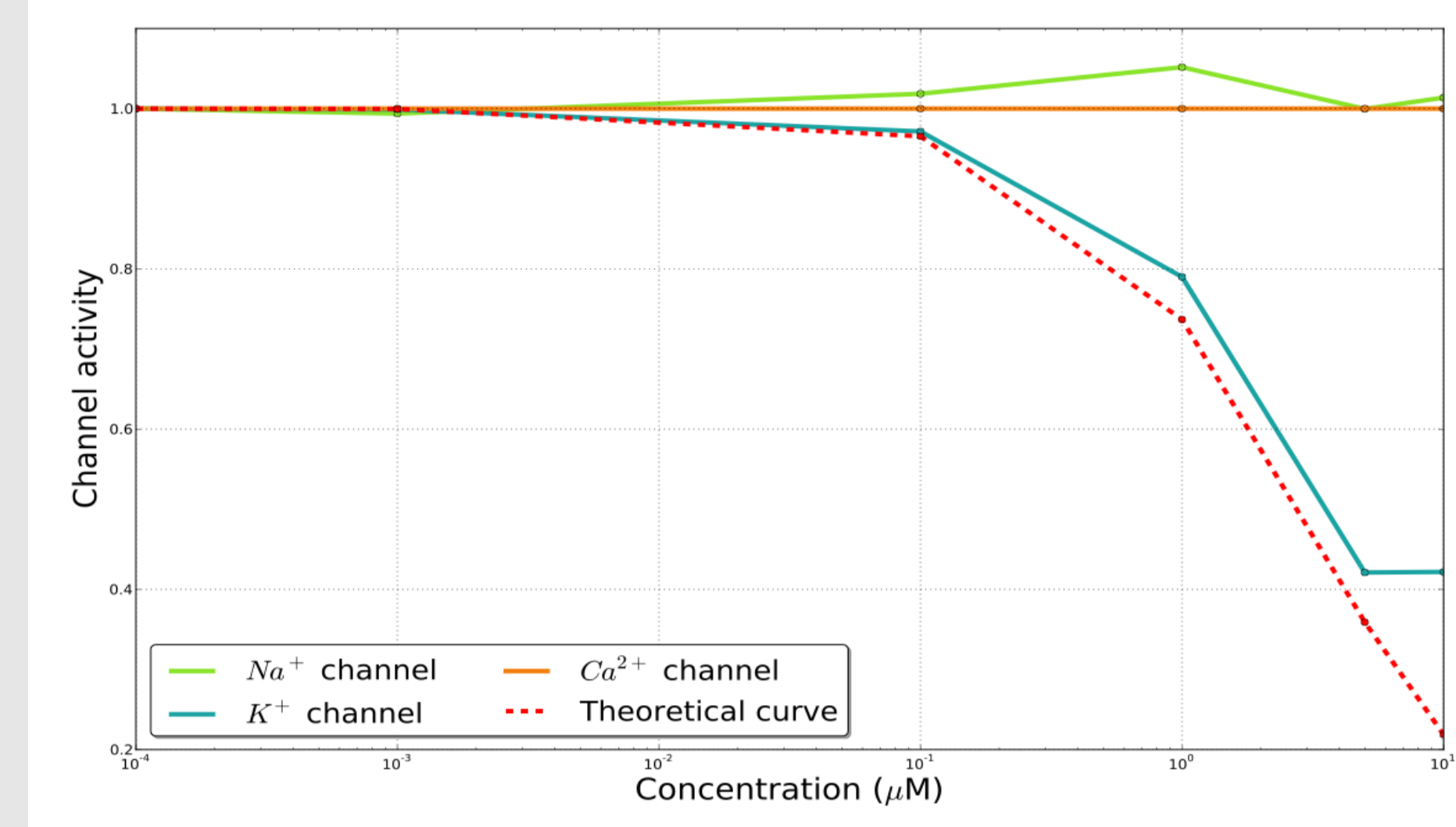


## Inverse problem: channel activity identification

- IC50 estimation of real data with the inverse problem approach**

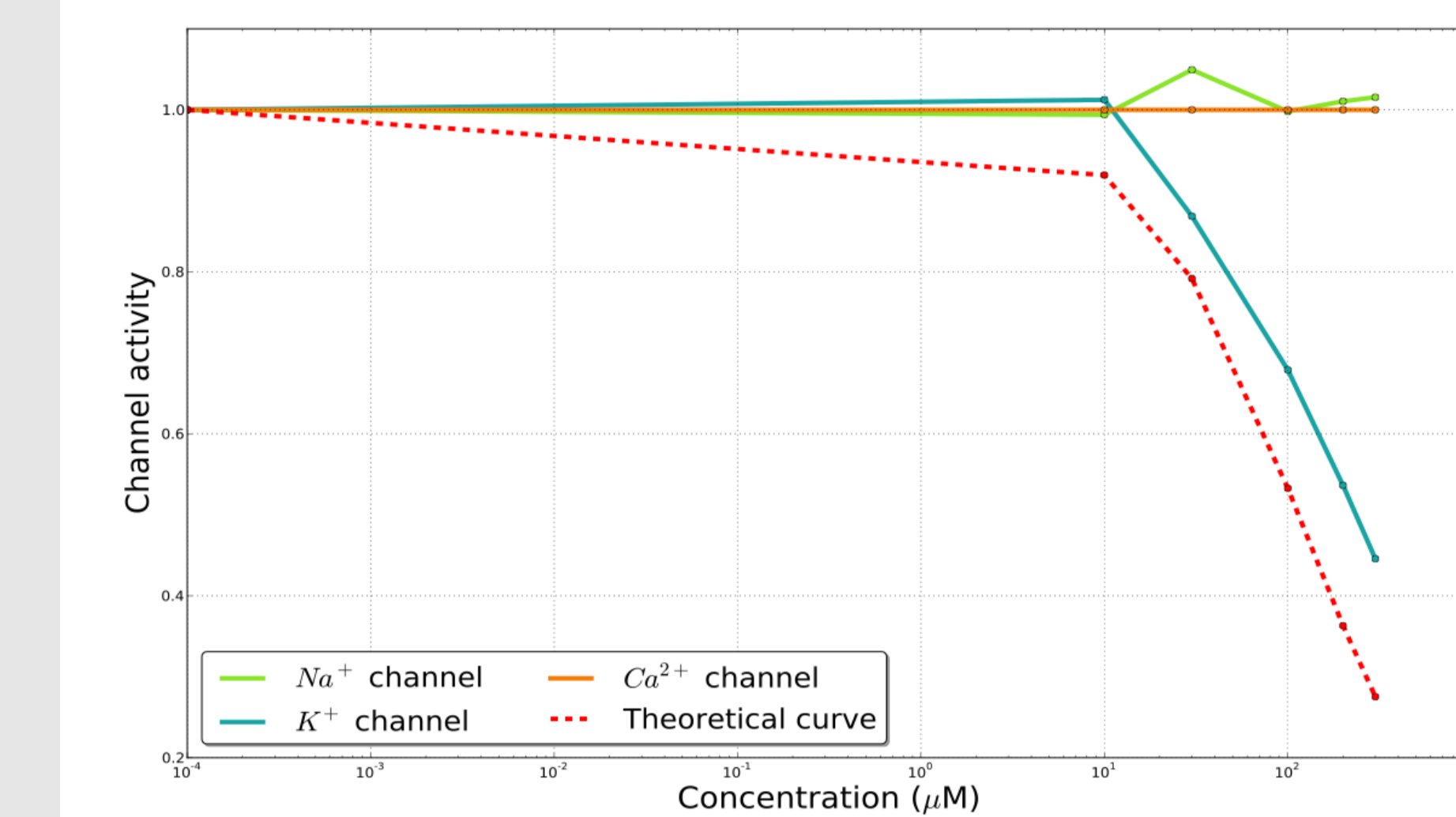
- Model: MV.
- Data: real experiments (CDI) processed with NOTOCORD-fps.
- Identification of 3 channel conductances: fast inward, slow inward and outward currents (denoted "Na+", "Ca2+", "K+" for simplicity).
- Theoretical curves obtained with the published IC50 and the function:  $f(c) = \left(1 + \frac{c}{IC_{50}}\right)^{-1}$

Channel activity identification using Ivabradine as observation



Ivabradine  
 $IC_{50}$  identified *in silico*  $\approx 3.7 \mu M$   
 Theoretical curve with  $IC_{50}(I_{Kf}) = 2.8 \mu M$   
 (Shattock et al. - British J Cardiology, 2006)

Channel activity identification using Moxifloxacin as observation

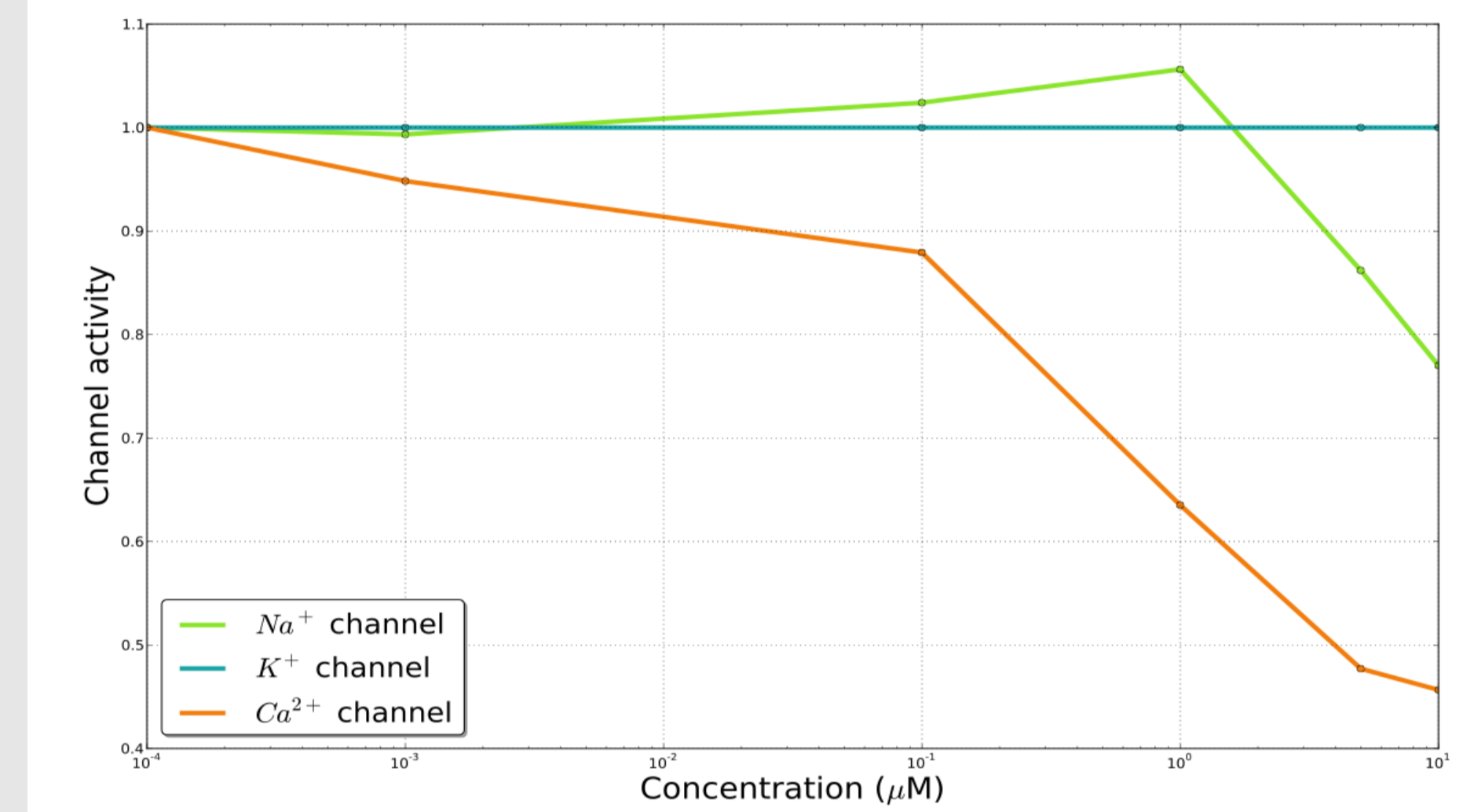


Moxifloxacin  
 $IC_{50}$  identified *in silico*  $\approx 220 \mu M$   
 Theoretical curve with  $IC_{50}(I_{Kr}) = 114 \mu M$   
 (Alexandrou et al., British J Pharmacol. 2006)

→ realistic values of IC50 found by the inverse problem approach.

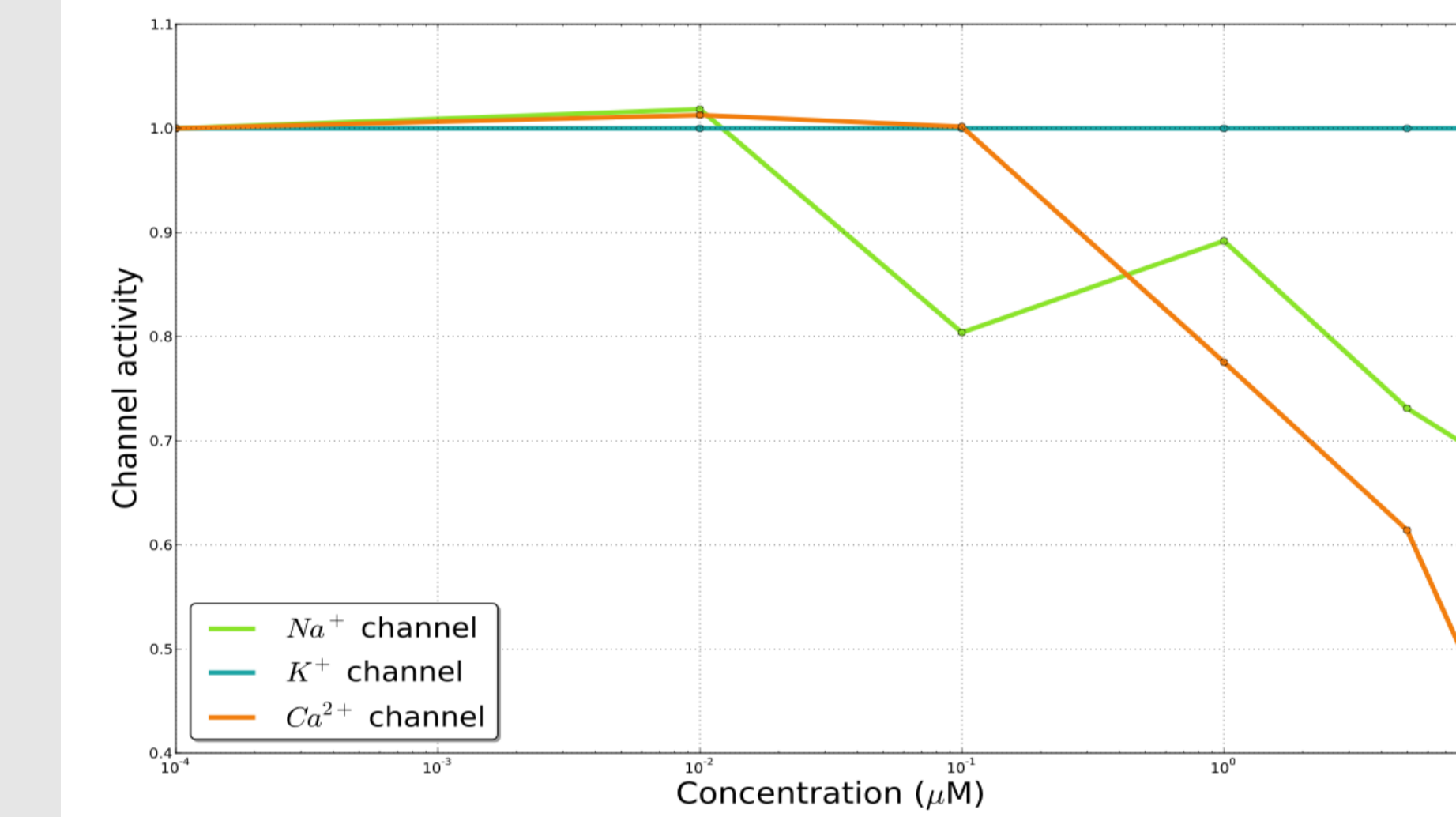
- Results with other compounds:

Channel activity identification using Diltiazem as observation



Diltiazem

Channel activity identification using SEA0400 as observation



SEA0400

## Classification

- Training with real experimental data**

Classification probabilities obtained with SVM using experimental data\* for training

Means (8 electrodes tested per drug)

	Ivabradine	Mexiletine	Moxifloxacin	Diltiazem	JNJ303	Dofetilide	BayK**	SEA0400	Ranolazine	Nimodipine
Na antagonist	0.0	<b>0.98</b>	0.04	<b>0.48</b>	0.16	0.0	0.0	0.04	<b>0.38</b>	0.03
K antagonist	<b>1.0</b>	0.0	0.0	0.0	0.0	<b>1.0</b>	0.0	0.0	0.0	0.0
Ca antagonist	0.0	0.01	<b>0.66</b>	<b>0.43</b>	0.41	0.0	0.1	<b>0.86</b>	0.33	<b>0.92</b>
Ca agonist	0.0	0.01	0.3	0.08	<b>0.43</b>	0.0	<b>0.9</b>	0.1	0.29	0.05

\*Ivabradine, Mexiletine, BayK and SEA0400 (as a Calcium blocker). \*\*4 electrodes tested for the prediction.

→ Moxifloxacin, Diltiazem, JNJ303 are not correctly classified.

- Training with 4096 in silico experiments (results of simulation of a simplified model)**

Classification probabilities obtained with SVM using MV model (4K samples) for training

Means (8 electrodes tested per drug)

	Ivabradine	Mexiletine	Moxifloxacin	Diltiazem	JNJ303	Dofetilide	BayK*	SEA0400	Ranolazine	Nimodipine
Na antagonist	0.0	<b>0.99</b>	0.0	0.01	0.22	0.0	0.1	0.0	<b>0.39</b>	0.0
Na agonist	0.05	0.0	0.14	0.0	0.0	0.0	0.12	0.0	0.0	0.01
K antagonist	0.0	0.0	<b>0.8</b>	0.0	<b>0.77</b>	0.0	0.0	0.0	<b>0.61</b>	0.0
K agonist	0.0	0.0	0.0	0.22	0.0	0.0	<b>0.39</b>	0.42	0.0	<b>0.68</b>
Ca antagonist	0.0	0.0	0.01	<b>0.77</b>	0.0	0.0	0.0	<b>0.58</b>	0.0	<b>0.31</b>
Ca agonist	<b>0.95</b>	0.0	0.05	0.0	0.01	<b>1.0</b>	<b>0.39</b>	0.0	0.0	0.0

\*4 electrodes tested for the prediction.

→ Ivabradine, Dofetilide, Ranolazine, Nimodipine are not correctly classified.

- Training with the real experimental data and the 4096 in silico experiments**

Classification probabilities obtained with SVM using MV model (4K samples) and experimental data\* for training

Means (8 electrodes tested per drug)

	Ivabradine	Mexiletine	Moxifloxacin	Diltiazem	JNJ303	Dofetilide	BayK**	SEA0400	Ranolazine	Nimodipine
Na antagonist	0.0	<b>0.98</b>	0.0	0.0	0.19	0.0	0.01	0.0	<b>0.31</b>	0.01
Na agonist	0.0	0.0	0.16	0.0	0.0	0.0	0.0	0.0	0.0	0.0
K antagonist	<b>0.83</b>	0.0	<b>0.6</b>	0.0	<b>0.73</b>	<b>0.98</b>	0.0	0.0	0.48	0.0
K agonist	0.0	0.0	0.0	0.12	0.0	0.0	0.0	0.45	0.0	0.48
Ca antagonist	0.0	0.03	0.05	<b>0.87</b>	0.01	0.01	0.0	<b>0.54</b>	0.01	<b>0.51</b>
Ca agonist	0.17	0.0	0.19	0.0	0.06	0.01	<b>0.99</b>	0.0	0.0	0.0

\*Ivabradine, BayK and SEA0400 (as a Calcium blocker). \*\*4 electrodes tested for the prediction.

→ By mixing real and *in silico* data for the training, all the compounds are correctly classified.

## Conclusion

- We proposed a mathematical model of MEA (direct and inverse problems) and we trained a machine learning algorithm with real and in silico signals.
- Direct problem: allow to reproduce *in silico* observed phenomena and better understand some features of the signals (EAD, ...).
- Inverse problem: identify the channel activity from synthetic and real signals, determine IC50.
- Limitations and future work:
  - The cell model used for the inverse problem should be replaced by a more comprehensive one.
  - To improve inverse problems and classification, additional biomarkers should be identified on the FP.
  - Machine learning algorithm has to be trained with more experimental and synthetic signals.
  - The overall methodology has to be tested with more compounds.